

Journal Pre-proof

The effects of Curcumin supplementation on oxidative stress, Sirtuin-1 and Peroxisome proliferator activated receptor γ coactivator 1 α gene expression in polycystic ovarian syndrome (PCOS) patients: a randomized placebo-controlled clinical trial

Javad Heshmati, Fereshteh Golab, Mojgan Morvaridzadeh, Eric Potter, Maryam Akbari-Fakhrabadi, Farnaz Farsi, Sara Tanbakooei, Farzad Shidfar



PII: S1871-4021(20)30003-5

DOI: <https://doi.org/10.1016/j.dsx.2020.01.002>

Reference: DSX 1556

To appear in: *Diabetes & Metabolic Syndrome: Clinical Research & Reviews*

Received Date: 29 December 2019

Revised Date: 5 January 2020

Accepted Date: 6 January 2020

Please cite this article as: Heshmati J, Golab F, Morvaridzadeh M, Potter E, Akbari-Fakhrabadi M, Farsi F, Tanbakooei S, Shidfar F, The effects of Curcumin supplementation on oxidative stress, Sirtuin-1 and Peroxisome proliferator activated receptor γ coactivator 1 α gene expression in polycystic ovarian syndrome (PCOS) patients: a randomized placebo-controlled clinical trial, *Diabetes & Metabolic Syndrome: Clinical Research & Reviews*, <https://doi.org/10.1016/j.dsx.2020.01.002>.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2020 Published by Elsevier Ltd on behalf of Diabetes India.

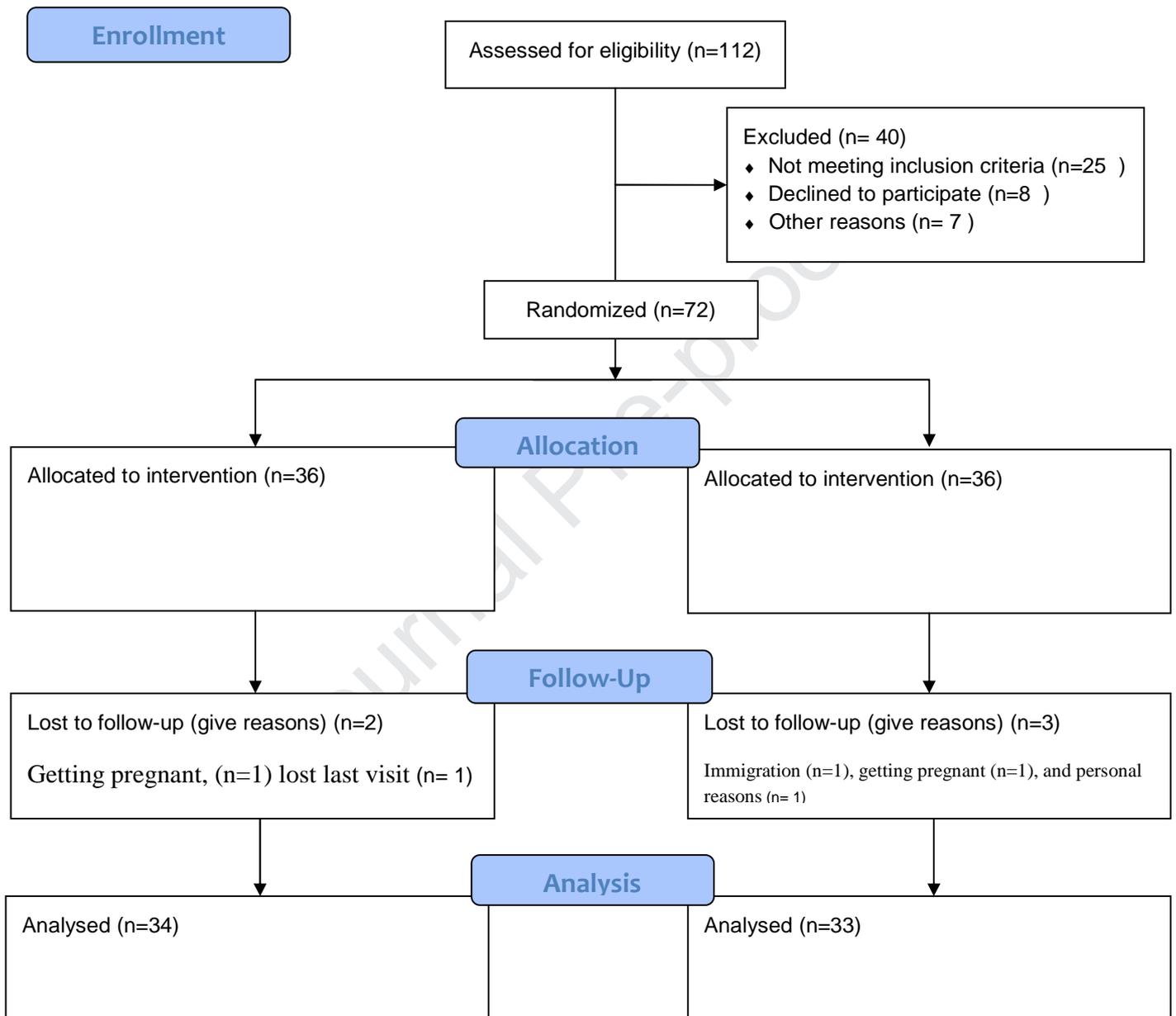


Figure 1. Flow diagram of the study

1 **The effects of Curcumin supplementation on oxidative stress,**
2 **Sirtuin-1 and Peroxisome proliferator activated receptor γ**
3 **coactivator 1 α gene expression in polycystic ovarian syndrome**
4 **(PCOS) patients: a randomized placebo-controlled clinical trial**

5 **Javad Heshmati¹, Fereshteh Golab², Mojgan Morvaridzadeh³, Eric Potter⁴, Maryam**
6 **Akbari-Fakhrabadi¹, Farnaz Farsi¹, Sara Tanbakooei², Farzad Shidfar^{1*}**

7 1. Department of Nutrition, School of Public Health, Iran University of Medical Sciences,
8 Tehran, Iran.

9 2. Cellular and Molecular Research Center, Iran University of Medical Sciences, Tehran,
10 Iran

11 3. Department of Nutritional Science, School of Nutritional Science and Food Technology,
12 Kermanshah University of Medical Sciences, Kermanshah, Iran

13 4. Baylor Scott & White Research Institute, Dallas, Texas, USA E-mail: Eric.potter@bswhealth.org

14
15 ***Farzad Shidfar (corresponding author)**

16 Department of Nutrition, School of Public Health, Iran University of Medical Sciences, Tehran,
17 Iran, Shahid Hemmat Highway, Tehran, 1449614535, IRAN P.O Box: 14665-354

18 **E-mail: shidfar.f@iums.ac.ir**

19 **Telephone: +9821-88622755**

20 **Funding Sources**

21 None.

22 **Conflicts of Interest**

23 The authors have no conflicts of interest to declare.
24
25

Highlights

- Polycystic ovary syndrome (PCOS) is a common gynecological disease
- Oxidative stress and imbalance between oxidants and antioxidants correlate with PCOS complications
- Curcumin is a potent antioxidant in turmeric and it has been determined to impact the expression of antioxidant related genes.
- Curcumin can improve body antioxidant enzymes by impacting related gene

expression in patients with PCOS

Journal Pre-proof

26 **Abstract**

27 **Background & Aims:** Curcumin is a biologically active phytochemical ingredient found in
28 turmeric and has antioxidant pharmacologic actions that may benefit patients with polycystic
29 ovarian syndrome (PCOS). The aim in this trial was to evaluate the efficacy of curcumin
30 supplementation on oxidative stress enzymes, sirtuin-1 (SIRT1) and Peroxisome proliferator
31 activated receptor γ coactivator 1 α (PGC1 α) gene expression in PCOS patients.

32 **Methods:** Seventy-two patients with PCOS were recruited for this randomized, double-blinded,
33 clinical trial. Thirty-six patients received curcumin, 1500 mg (three times per day), and 36
34 patients received placebo for 3 months. Gene expression of SIRT1, PGC1 α and serum activity of
35 glutathione peroxidase (Gpx) and superoxide dismutase (SOD) enzymes were evaluated at the
36 beginning of trial and at 3-month follow-up.

37 **Results:** Sixty-seven patients with PCOS completed the trial. Curcumin supplementation
38 significantly increased gene expression of PGC1 α ($p=0.011$) and activity of the Gpx enzyme
39 ($p=0.045$). Curcumin also non-significantly increased gene expression of SIRT1 and activity of
40 the SOD enzyme.

41 **Conclusions:** Curcumin seems to be an efficient reducer of oxidative stress related complications
42 in patients with PCOS. Further studies on curcumin should strengthen our findings.

43 **Keywords:** Curcumin; Glutathione peroxidase; PGC1 α ; SIRT1; Superoxide dismutase

44

45

46

47

48

49 **Background**

50 Polycystic ovarian syndrome (PCOS) is considered to be the most common world-wide cause of
51 infertility [1]. PCOS is also one of the major metabolic disorders in women of developing
52 countries[2, 3]. Inflammation and oxidative stress may cause PCOS and related
53 complications[4]. Oxidative stress refers to the imbalance between excessive production of
54 reactive oxygen species (ROS) and a limited amount of body antioxidant defense [5]. Various
55 factors are involved in oxidative stress control. The first defensive line against ROS are
56 detoxifying enzymes like superoxide dismutase (SOD) and glutathione peroxidase (GPx). These
57 enzymes are found in the mitochondria, cytoplasm and cell peroxisomes[6]. The next line of
58 antioxidant defense is the limitation of ROS production by uncoupling proteins (UCPs)[7]. These
59 UCPs reduce the production of ROS by decreasing the electrochemical potential within the inner
60 layer of the mitochondria which shortens most of the steps in the electron transfer chain[8].

61 Peroxisome proliferator activated receptor γ coactivator-1 α (PGC-1 α), is an activator in the cell
62 nucleus that regulates various biological actions, including: mitochondrial biogenesis; fatty acid
63 oxidation and glucose metabolism. PGC-1 α also has several roles in the metabolism of fatty
64 acids and insulin sensitivity[9]. PGC-1 α has been demonstrated to be an effective factor for
65 increasing levels of SOD and Gpx enzymes and enhancing the antioxidant defense[10]. PGC-1 α
66 is mainly expressed in brown adipose tissue (BAT), and most other cells in the body, and is
67 responsible for inducing the differentiation of BAT. The expression of this gene has been shown
68 to increase the expression of UCP-1, which is the main thermogenesis inducer factor in
69 BAT[11]. Studies in recent years have also demonstrated polymorphism of the PCG-1a gene in
70 women with PCOS is different compared to healthy controls[12]. Other studies have shown that

71 PGC-1 α gene expression alters the insulin resistance in PCOS[13]. It seems that increasing PGC-
72 1 α expression by increasing the expression of UCP-1 can elevate the oxidation of fats, induce
73 thermogenesis, reduce lipogenesis, and decrease obesity in PCOS women.

74 Silent Information Regulator 1 (Sirt1) is also a NAD⁺ dependent histone deacetylase in the
75 pathway of insulin secretion. Sirt1 is involved in the expression of antioxidant defense factors
76 and enzymes and also plays a role in apoptosis[14]. Sirt1 can regulate oxidative stress and
77 prevent damage to DNA through its impact on the p53 protein[15]. Sirt1 contributes to the
78 deacetylation of the PGC-1 α gene, thereby increasing the rate of thermogenesis and oxidation of
79 lipids by activating Peroxisome proliferator-activated receptor- α (PPAR- α). The effects of Sirt1
80 on glucose and insulin hemostasis are also applied through PGC-1 α [16].

81 Curcumin is a naturally active phytochemical derived from the traditional spice turmeric[17].
82 Curcumin has demonstrated anti-inflammatory and anti-oxidant activity through multiple
83 mechanisms such as its impact on gene expression and cellular signaling [18, 19]. The
84 antioxidant effects of curcumin are represented through increasing gene expression of SOD and
85 GPx[20]. Curcumin has been shown to reduce blood glucose by inhibiting liver gluconeogenesis
86 through affecting the 5' AMP-activated protein kinase (AMPK) signaling pathway[21]. The
87 effect of curcumin on PGC-1 α has been evaluated only in cellular studies in which curcumin has
88 been shown to increase the expression of PGC-1 α through its effect on AMPK[22].

89 Our hypothesis is that curcumin can help to improve the complications of PCOS through
90 regulating the gene expression of antioxidant enzymes. Furthermore, only a limited number of
91 cellular studies have investigated the effect of curcumin, or similar polyphenols, on Sirt1 gene
92 expression. This study aims to investigate the effect of curcumin supplementation on the
93 expression of PGC-1 α and Sirt1 genes and oxidative factors in women with PCOS.

94

95 **Methods and Design**

96 Study Design

97 This was a randomized, double-blinded, clinical trial involving 72 overweight or obese [Body
98 mass index (BMI), $>25 \text{ kg/m}^2$] female patients with PCOS with impaired glucose tolerance at
99 our Arash hospital in Tehran, Iran. PCOS was diagnosed according to the Rotterdam criteria[23].
100 After providing informed consent, the subjects were studied for a period between January 2019
101 and June 2019. The study was approved by the Iranian Registry of Clinical Trials on 2019-01-23
102 and our registration reference is IRCT20091114002709N50 (<https://www.irct.ir/trial/35137>).
103 The subjects were divided into two groups (each group contain 36 subjects) by computer-based
104 block randomization. The intervention group received 1500 mg/day (500 mg Three times daily)
105 of curcumin, and the control group received 1500 mg/day of placebo (maltodextrin) for 12
106 weeks. Curcumin and placebo capsules were prepared by KAREN Pharma (Yazd, Iran), Shape,
107 size, smell and color of placebo capsules were completely similar to the curcumin capsules.
108 Personnel and patients was blinded to the treatment allocation. Height and waist circumference
109 were measured before treatment and at 3-month follow-up to the nearest 0.1 centimeter, and
110 bodyweight was measured to the nearest 100 grams. BMI was calculated as the weight (kg)
111 divided by the square of height (meters). All patients were advised not to change their physical
112 activity and dietary patterns during the intervention. Dietary intake was evaluated by 24h food
113 recall two times, in beginning and end of the intervention. Patient's compliance were evaluated
114 by weekly phone call of the researcher to each patients and monitoring for possible side effects
115 assessed as well.

116 In the beginning of the intervention and after 12 weeks of study, blood samples of 10 ml were
117 obtained after 12-14 hour of overnight fasting. Buffy coat of blood white cells was separated by
118 centrifugation of blood samples. RNA was extracted using RNX- plus Sinacolon Kit and then
119 cDNA was synthesized using SinaClon first strand cDNA synthesis kit. Real-time polymerase
120 chain reaction (PCR) was carried out based on the protocols described on Sinacolon kit
121 (SinaSYBR Blue HS-qPCRMix, 2x, Iran). GAPDH was used in real-time PCR as the
122 housekeeping gene. The primer sequences that we used in real-time PCR are described in **Table**
123 **1**. Serum concentration of GPX and SOD activity were evaluated by the methods prescribed by
124 Paglia et al. and Sun et al., respectively[24, 25].

125 Statistical analysis was performed using SPSS Software v.21. Data were shown as mean \pm SE.
126 Baseline characteristics were compared among the two intervention groups using an independent
127 sample t-test for continuous data and a chi-square test for ordinal data. The magnitude of the
128 effect is presented as mean difference and its 95% confidence interval. Final antioxidant
129 enzymes and gene expression results was adjusted for possible confounders such as basic value
130 of dependent variable, treatment type, age and BMI.

131

132 **Results**

133 One hundred and twelve women with PCOS were initially consented to participate in the trial, of
134 which 40 subjects did not meet the inclusion criteria. Seventy-two patients met inclusion criteria
135 and were included in this study. Five patients stopped the supplement intake due to personal
136 reasons, pregnancy, or immigration and were excluded from the trial. Therefore, at the end of
137 treatment, 67 patients completed the study. **Figure 1** presents a flow diagram of the study.

138 There were no statistically significant differences between the mean values of participant's age at
139 the beginning of the trial. **Table 2** presents the baseline and endpoints of anthropometric
140 parameters of the participants. There were no significant differences between the anthropometric
141 indices within the various groups at the beginning of the treatment. Neither curcumin nor
142 placebo had significant impacts on anthropometric parameters. **Table 3** shows the dietary intakes
143 of the trial groups at the beginning and end of the intervention according on their 24-hr recall
144 analyses. As presented, there were no significant differences in energy, macronutrients, or main
145 antioxidants of the diet between the study groups at the baseline and end-point of the
146 intervention.

147 *Serum antioxidant enzymes*

148 As indicated in **Table 4**, curcumin supplementations significantly increased serum activity of
149 GPx ($p=0.041$). This effect was still significant after adjustment for age, BMI and baseline
150 values of GPx ($p=0.010$). Curcumin supplementation increased serum activity of SOD, but it
151 was not statistically significant ($p= 0.075$), this impact also did not change after adjustment
152 ($p=0.0730$).

153 *Gene expression findings*

154 **Figures 2 and 3** show the effect of curcumin on gene expression of PGC-1a and SIRT1
155 respectively. Outcomes of this trial indicated that after adjustment for confounders like age, BMI
156 and baseline values, the gene expressions of PGC-1a according to $2^{-\Delta\Delta Ct}$ calculation were
157 statistically increased in the curcumin group compared to the placebo ($p =0.031$). These results
158 also indicated that curcumin supplementation compared to placebo increased SIRT1 gene
159 expression, but after adjustment, this increase was not statistically significant (**Table 5**).

160

161 **Discussion**

162 This is the first randomized clinical trial to evaluate the impact of curcumin on gene expression
163 of SIRT1 and PGC-1 α in humans. We have demonstrated that curcumin, administered to women
164 with PCOS, led to increased GPx and gene expression of PGC-1 α , as well as a non-significant
165 increase in SOD and gene expression of SIRT1, after 3 months compared to placebo. Curcumin
166 supplementation in 1500 mg/d did not show any adverse events for participants and according to
167 previous studies this dose is safe and effective in this regards [26, 27].

168 The pathogenesis of PCOS still remains unclear. Multiple features and complications of PCOS,
169 including excessive androgens, insulin resistance, and abdominal adiposity, may lead to increase
170 local and systemic oxidative stress [28-30]. Reciprocally, oxidative stress also may worsen
171 these metabolic abnormalities. Reducing oxidative stress in PCOS patients, or strengthening their
172 antioxidant defense systems, can be helpful in decreasing and improving the complications of the
173 disease[31].

174 PGC-1 α expression increases antioxidant enzymes such as Gpx[32], and reduces oxidative stress.
175 PGC-1 α can also potentially decrease production of mitochondrial-driven ROS[33]. This
176 randomized clinical trial has demonstrated that curcumin significantly increases gene expression
177 of PGC-1 α and Gpx activity. This likely occurs because the curcumin impacts Gpx activity
178 through increased gene expression of PGC-1 α . Previous cellular studies have indicated that
179 curcumin stimulation increased gene expression of PGC-1 α , and the impacts of curcumin on
180 PGC-1 α expression were associated with the activation of adenosine monophosphate-activated
181 protein kinase (AMPK)[22]. It has been shown that curcumin also increased antioxidant enzymes

182 such as SOD transcriptions and activity by the AMPK/PGC-1 α axis[34]. It has been
183 demonstrated in several in vitro studies that curcumin performs its antioxidant actions through
184 changes in several nuclear factors such as Nrf2, PPAR γ , and NF- κ B[35-37]. It has also been
185 shown that these factors are regulated by the AMPK/PGC-1 α axis[38]. PGC-1 α enhancement
186 also may have other beneficial effects on PCOS such as increased lipid oxidation through
187 activation of PPAR- α [39]. Abdominal obesity is one of the major risk factors for PCOS, and
188 PPAR- α is one of the main factors to increase fat oxidation and thermogenesis of fat tissues[40].
189 So it seems that curcumin may have beneficial effects to reduce fat mass and obesity
190 complications in patients with PCOS through increased gene expression of PGC-1 α and so
191 PPAR- α .

192 In our study, curcumin supplementation non-significantly increased Sirt1 gene expression.
193 Previous in vivo and in vitro studies indicated that curcumin treatment improved mitochondrial
194 oxidative damage through the activation of SIRT1 signaling[41, 42]. In vitro studies also
195 indicated that curcumin supplementation can attenuate down-regulation of SIRT1, which showed
196 that the activation of SIRT1 might be due to the protective effect of curcumin[43]. Perhaps the
197 low dose and short duration of our study due to human research considerations have led to the
198 inability to draw significant results in this area. However, Sirt1 is in the signaling pathway of the
199 AMPK/PGC-1 α axis[44], and according to our results, curcumin can increase PGC-1 α gene
200 expression and this may indicate that the increase in Sirt1 was associated with an increase in
201 PGC-1 α . It has been suggested that SIRT1-mediated deacetylation of PGC-1 α is attributed to the
202 anti-oxidant activity of curcumin [45]. Insulin resistance is one of the main complications of
203 PCOS. Growing evidence proposes that SIRT1 regulates glucose and insulin metabolism through
204 its deacetylase function for several known substrates, and also it has been shown that SIRT1 has

205 a positive activity in the metabolic pathway by its direct or indirect impact on insulin
206 signaling[46]. There is also evidence that SIRT1 upregulation induces a glucose-dependent
207 insulin secretion from pancreatic β cells[47], and directly induces insulin sensitivity[48]. This is
208 the first randomized clinical trial which evaluated the effect of curcumin on gene expression in
209 human model, however this study also have some limitations. For example maybe higher dose,
210 longer duration of supplementation could make us to draw more resolute conclusion in this
211 regards.

212 In conclusion, the results of this randomized clinical trial indicate that curcumin supplementation
213 significantly increases PGC-1 α gene expression and serum enzyme activity of Gpx. We have
214 also demonstrated that curcumin supplementation increases SIRT1 gene expression and SOD
215 enzyme activity in non-significant manner. These results indicate that curcumin is potentially a
216 supplementary medication for the management of PCOS. However, there is still a need for larger
217 and longer randomized clinical trials on other clinical factors to draw a clear conclusion about
218 the effect of curcumin on PCOS.

219

220 **References**

- 221 1. *Health and fertility in World Health Organization group 2 anovulatory women*. Human
222 Reproduction Update, 2012. **18**(5): p. 586-599.
- 223 2. Ranasinghe, P., et al., *Prevalence and trends of metabolic syndrome among adults in the asia-*
224 *pacific region: a systematic review*. BMC public health, 2017. **17**(1): p. 101.
- 225 3. Misra, A. and L. Khurana, *Obesity and the metabolic syndrome in developing countries*. The
226 Journal of Clinical Endocrinology & Metabolism, 2008. **93**(11_supplement_1): p. s9-s30.
- 227 4. Sabuncu, T., et al., *Oxidative stress in polycystic ovary syndrome and its contribution to the risk*
228 *of cardiovascular disease* ☆. Clinical biochemistry, 2001. **34**(5): p. 407-413.
- 229 5. Rezaie, A., R.D. Parker, and M. Abdollahi, *Oxidative stress and pathogenesis of inflammatory*
230 *bowel disease: an epiphenomenon or the cause?* Digestive diseases and sciences, 2007. **52**(9): p.
231 2015-2021.
- 232 6. Vats, P., et al., *Association of superoxide dismutases (SOD1 and SOD2) and glutathione*
233 *peroxidase 1 (GPx1) gene polymorphisms with type 2 diabetes mellitus*. Free radical research,
234 2015. **49**(1): p. 17-24.

- 235 7. Phulukdaree, A., et al., *Uncoupling protein 2– 866G/A and uncoupling protein 3– 55C/T*
 236 *polymorphisms in young South African Indian coronary artery disease patients*. *Gene*, 2013.
 237 **524**(2): p. 79-83.
- 238 8. Li, F., et al., *Yiqihuoxue decoction protects against post-myocardial infarction injury via*
 239 *activation of cardiomyocytes PGC-1 α expression*. *BMC complementary and alternative medicine*,
 240 2018. **18**(1): p. 253.
- 241 9. Mirzaei, K., et al., *Insulin resistance via modification of PGC1 α function identifying a possible*
 242 *preventive role of vitamin D analogues in chronic inflammatory state of obesity. A double blind*
 243 *clinical trial study*. *Minerva medica*, 2014. **105**(1): p. 63-78.
- 244 10. Roy, V.K., R. Verma, and A. Krishna, *Carnitine-mediated antioxidant enzyme activity and Bcl2*
 245 *expression involves peroxisome proliferator-activated receptor- γ coactivator-1 α in mouse testis*.
 246 *Reproduction, Fertility and Development*, 2017. **29**(6): p. 1057-1063.
- 247 11. Gallardo-Montejano, V.I., et al., *Nuclear Perilipin 5 integrates lipid droplet lipolysis with PGC-*
 248 *1 α /SIRT1-dependent transcriptional regulation of mitochondrial function*. *Nature*
 249 *communications*, 2016. **7**: p. 12723.
- 250 12. Reddy, T.V., et al., *Polymorphisms in the TFAM and PGC1- α genes and their association with*
 251 *polycystic ovary syndrome among South Indian women*. *Gene*, 2018. **641**: p. 129-136.
- 252 13. Chen, L., et al., *Explore the Relationship between Insulin Resistance and PGC1 α in PCOS Mice*.
 253 *Open Journal of Endocrine and Metabolic Diseases*, 2018. **8**(03): p. 71.
- 254 14. Howitz, K.T., et al., *Small molecule activators of sirtuins extend *Saccharomyces cerevisiae**
 255 *lifespan*. *Nature*, 2003. **425**(6954): p. 191.
- 256 15. Luo, J., et al., *Acetylation of p53 augments its site-specific DNA binding both in vitro and in vivo*.
 257 *Proceedings of the National Academy of Sciences of the United States of America*, 2004. **101**(8):
 258 p. 2259-2264.
- 259 16. Tao, X., et al., *Regulatory effects of the AMPK α -SIRT1 molecular pathway on insulin resistance in*
 260 *PCOS mice: An in vitro and in vivo study*. *Biochemical and biophysical research communications*,
 261 2017. **494**(3): p. 615-620.
- 262 17. Gupta, S.C., G. Kismali, and B.B. Aggarwal, *Curcumin, a component of turmeric: from farm to*
 263 *pharmacy*. *Biofactors*, 2013. **39**(1): p. 2-13.
- 264 18. Kunnumakkara, A.B., et al., *Curcumin, the golden nutraceutical: multitargeting for multiple*
 265 *chronic diseases*. *British journal of pharmacology*, 2017. **174**(11): p. 1325-1348.
- 266 19. Ghorbani, M., et al., *Curcumin-lipoic acid conjugate as a promising anticancer agent on the*
 267 *surface of gold-iron oxide nanocomposites: A pH-sensitive targeted drug delivery system for*
 268 *brain cancer theranostics*. *European Journal of Pharmaceutical Sciences*, 2018. **114**: p. 175-188.
- 269 20. Fuentes, F., et al., *Nrf2-mediated antioxidant and detoxifying enzyme induction by a*
 270 *combination of curcumin and sulforaphane*. *gene expression*, 2016. **11**: p. 18.
- 271 21. Kim, J.H., et al., *Curcumin stimulates glucose uptake through AMPK-p38 MAPK pathways in L6*
 272 *myotube cells*. *Journal of cellular physiology*, 2010. **223**(3): p. 771-778.
- 273 22. Zhai, X., et al., *Curcumin regulates peroxisome proliferator-activated receptor- γ coactivator-1 α*
 274 *expression by AMPK pathway in hepatic stellate cells in vitro*. *European journal of pharmacology*,
 275 2015. **746**: p. 56-62.
- 276 23. Franks, S., *Diagnosis of polycystic ovarian syndrome: in defense of the Rotterdam criteria*. *The*
 277 *Journal of Clinical Endocrinology & Metabolism*, 2006. **91**(3): p. 786-789.
- 278 24. Paglia, D.E. and W.N. Valentine, *Studies on the quantitative and qualitative characterization of*
 279 *erythrocyte glutathione peroxidase*. *The Journal of laboratory and clinical medicine*, 1967. **70**(1):
 280 p. 158-169.
- 281 25. Sun, Y., L.W. Oberley, and Y. Li, *A simple method for clinical assay of superoxide dismutase*.
 282 *Clinical chemistry*, 1988. **34**(3): p. 497-500.

- 283 26. Faghfoori, Z., et al., *Nutritional management in women with polycystic ovary syndrome: A review*
284 *study*. Diabetes & Metabolic Syndrome: Clinical Research & Reviews, 2017. **11**: p. S429-S432.
- 285 27. Dehghani, S., et al., *Multifunctional MIL-Cur@ FC as a theranostic agent for magnetic resonance*
286 *imaging and targeting drug delivery: in vitro and in vivo study*. Journal of Drug Targeting,
287 2019(just-accepted): p. 1-37.
- 288 28. Liu, S., G. Navarro, and F. Mauvais-Jarvis, *Androgen excess produces systemic oxidative stress*
289 *and predisposes to β -cell failure in female mice*. PLoS one, 2010. **5**(6): p. e11302.
- 290 29. Vincent, H.K. and A.G. Taylor, *Biomarkers and potential mechanisms of obesity-induced oxidant*
291 *stress in humans*. International journal of obesity, 2006. **30**(3): p. 400.
- 292 30. Fazelian, S., et al., *Chromium supplementation and polycystic ovary syndrome: A systematic*
293 *review and meta-analysis*. Journal of Trace Elements in Medicine and Biology, 2017. **42**: p. 92-
294 96.
- 295 31. Amini, L., et al., *Antioxidants and management of polycystic ovary syndrome in Iran: A*
296 *systematic review of clinical trials*. Iranian journal of reproductive medicine, 2015. **13**(1): p. 1.
- 297 32. Saboori, S., et al., *Beneficial effects of omega-3 and vitamin E coadministration on gene*
298 *expression of SIRT1 and PGC1 α and serum antioxidant enzymes in patients with coronary artery*
299 *disease*. Nutrition, Metabolism and Cardiovascular Diseases, 2016. **26**(6): p. 489-494.
- 300 33. Vazquez, F., et al., *PGC1 α expression defines a subset of human melanoma tumors with*
301 *increased mitochondrial capacity and resistance to oxidative stress*. Cancer cell, 2013. **23**(3): p.
302 287-301.
- 303 34. El-Bahr, S., *Effect of curcumin on hepatic antioxidant enzymes activities and gene expressions in*
304 *rats intoxicated with aflatoxin B1*. Phytotherapy research, 2015. **29**(1): p. 134-140.
- 305 35. Baliga, M.S., et al., *Curcumin, an active component of turmeric in the prevention and treatment*
306 *of ulcerative colitis: preclinical and clinical observations*. Food & function, 2012. **3**(11): p. 1109-
307 1117.
- 308 36. Ghorbani, Z., A. Hekmatdoost, and P. Mirmiran, *Anti-hyperglycemic and insulin sensitizer effects*
309 *of turmeric and its principle constituent curcumin*. International journal of endocrinology and
310 metabolism, 2014. **12**(4).
- 311 37. Ghosh, S.S., T.W. Gehr, and S. Ghosh, *Curcumin and chronic kidney disease (CKD): major mode of*
312 *action through stimulating endogenous intestinal alkaline phosphatase*. Molecules, 2014.
313 **19**(12): p. 20139-20156.
- 314 38. Amel Zabihi, N., et al., *Is there a role for curcumin supplementation in the treatment of non-*
315 *alcoholic fatty liver disease? The data suggest yes*. Current pharmaceutical design, 2017. **23**(7):
316 p. 969-982.
- 317 39. Hondares, E., et al., *Peroxisome proliferator-activated receptor α (PPAR α) induces PPAR γ*
318 *coactivator 1 α (PGC-1 α) gene expression and contributes to thermogenic activation of brown fat*
319 *involvement of PRDM16*. Journal of Biological Chemistry, 2011. **286**(50): p. 43112-43122.
- 320 40. Gross, B., et al., *PPARs in obesity-induced T2DM, dyslipidaemia and NAFLD*. Nature Reviews
321 Endocrinology, 2017. **13**(1): p. 36.
- 322 41. Yang, Y., et al., *SIRT1 activation by curcumin pretreatment attenuates mitochondrial oxidative*
323 *damage induced by myocardial ischemia reperfusion injury*. Free Radical Biology and Medicine,
324 2013. **65**: p. 667-679.
- 325 42. Miao, Y., et al., *Curcumin pretreatment attenuates inflammation and mitochondrial dysfunction*
326 *in experimental stroke: The possible role of Sirt1 signaling*. Brain research bulletin, 2016. **121**: p.
327 9-15.
- 328 43. Xiao, J., et al., *Curcumin protects against myocardial infarction-induced cardiac fibrosis via SIRT1*
329 *activation in vivo and in vitro*. Drug design, development and therapy, 2016. **10**: p. 1267.

- 330 44. Vellinga, T.T., et al., *SIRT1/PGC1 α -dependent increase in oxidative phosphorylation supports*
331 *chemotherapy resistance of colon cancer*. *Clinical cancer research*, 2015. **21**(12): p. 2870-2879.
- 332 45. Jia, N., et al., *SIRT1-mediated deacetylation of PGC1 α attributes to the protection of curcumin*
333 *against glutamate excitotoxicity in cortical neurons*. *Biochemical and biophysical research*
334 *communications*, 2016. **478**(3): p. 1376-1381.
- 335 46. Cao, Y., et al., *SIRT1 and insulin resistance*. *Journal of Diabetes and its Complications*, 2016.
336 **30**(1): p. 178-183.
- 337 47. Bordone, L., et al., *Correction: SIRT1 regulates insulin secretion by repressing UCP2 in pancreatic*
338 *β cells*. *PLoS biology*, 2015. **13**(12): p. e1002346.
- 339 48. Hui, X., et al., *Adipocyte SIRT1 controls systemic insulin sensitivity by modulating macrophages in*
340 *adipose tissue*. *EMBO reports*, 2017. **18**(4): p. 645-657.

Table 1 Primers used in the current study.

PGC1-α -Forward	GTCAACATTCAAAGCAGCAGAGAG
PGC1-α-Reverse	GACACATAATCATTACCTACTGGAAGC
SIRT1- Forward	TAGTAGGCGGCTTGATGGTAATC
SIRT1- Reverse	GGTTCTTCTAAACTTGGACTCTGG

Table 2. Baseline characteristics after random assignment

		Total	Groups		P
			Intervention(A)(n=34)	Control(B)(n=3 3)	
Age	Median (range)	30 (18 to 52)	31 (18 to 41)	29 (20 to 52)	0.89 7 [†]
BMI	Median (range)	26.77 (17.75 to 39.56)	28.30 (19.05 to 39.56)	26.72 (17.75 to 36.00)	0. 230 [†]
Waist circumference	Median (range)	89.20 (61 to 117)	87.68 (61 to 117)	90.65 (70 to 114)	0.24 5 [†]

[†] Based on t-test.

BMI: Body mass index

Table 3. Dietary intake of study participants throughout the study

		Intervention (n=34)	Control (n=33)	95% CI		P value [†]
		Mean \pm SD	Mean \pm SD	Lower	Upper	
Energy intake (kcal/day)	Pre	2396.08 \pm 462.68	2361.93 \pm 534.52	-205.97	274.25	0.777
	Post	2226.16 \pm 443.30	2207.60 \pm 383.78	-178.73	215.84	0.852
	Change	-169.91 \pm 398.79	-154.33 \pm 516.96	-238.30	207.13	0.889
Carbohydrate intake (gr/day)	Pre	360.75 \pm 78.07	339.00 \pm 78.67	-15.71	59.21	0.251
	Post	348.40 \pm 87.89	328.02 \pm 88.91	-21.88	62.65	0.339
	Change	-12.34 \pm 62.76	-10.97 \pm 75.80	-34.84	32.11	0.935

Protein intake (gr/day)	Pre	92.27 ± 21.50	82.75 ± 28.14	-2.56	21.60	0.120
	Post	95.34 ± 24.56	85.94 ± 25.30	-2.52	22.33	0.114
	Change	3.07 ± 14.14	3.18 ± 15.93	-7.34	7.11	0.975
Vitamin E (mg/day)	Pre	11.34 ± 7.06	11.62 ± 9.06	-4.19	3.64	0.887
	Post	10.82 ± 5.83	9.61 ± 5.33	-1.45	3.87	0.368
	Change	-0.51 ± 6.21	-2.00 ± 8.10	-1.99	4.97	0.396
Vitamin C (mg/day)	Pre	108.55 ± 51.69	94.86 ± 59.16	-12.97	40.37	0.309
	Post	94.29 ± 57.16	90.97 ± 44.30	-20.94	27.59	0.785
	Change	-14.26 ± 41.35	-3.88 ± 55.49	-34.01	13.26	0.383

Data are presented as mean±SD

† Based on independent t-test.

*statistically significant

Table 4. Comparison of Gpx and SOD enzyme activities between the two groups

		INTERVENTION (N=34)	CONTROL (N=33)	95% CI		P VALUE†	ADJUSTED P VALUE§
GPX	Pre	108.83 ± 7.19	113.54 ± 9.48	-28.49	19.07	0.694	
	Post	141.66 ± 11.08	109.37 ± 7.54	5.49	59.10	0.019	
	Change	32.83± 13.10	-4.17 ± 11.99	1.56	72.44	0.041	0.010
	P# within	0.063	0.889				
SOD	Pre	202.77 ± 10.23	206.80 ± 12.16	-35.75	27.71	0.801	
	Post	241.02 ± 13.88	205.40 ± 16.01	-6.67	77.93	0.097	
	Change	38.25± 16.19	-1.40 ± 14.83	-4.17	83.47	0.075	0.073
	P# within	0.152	0.730				

Data are presented as mean±SE

† Based on independent t-test.

§ Based on linear regression (the included variables were: basic value of dependent variable, treatment type, age and BMI)

P value within groups based on paired t-test

*statistically significant

GPX: Glutathione peroxidase, SOD: Superoxide dismutase

Table 5. Comparison of Peroxisome proliferator activated receptor γ coactivator 1 α and Sirtuin-1 gene expression between the two groups

		Intervention (n=34)	Control (n=33)	95% CI		P value†	Adjusted P value§
Pgc1-α	Pre	1.123 \pm 0.277	1.168 \pm 0.140	-0.667	0.574	0.884	
	Post	2.686 \pm 0.353	1.717 \pm 0.281	0.072	1.867	0.034*	
	Change	1.563 \pm 0.494	0.548 \pm 0.299	-0.136	2.167	0.083	0.031*
	P [#] within	0.003*	0.074				
SIRT1	Pre	1.740 \pm 0.636	0.904 \pm 0.905	-1.483	3.154	0.549	
	Post	3.167 \pm 1.0578	1.050 \pm 0.826	-0.671	4.904	0.130	
	Change	1.427 \pm 0.547	0.146 \pm 0.145	0.061	2.499	0.048*	0.087
	P [#] within	0.023*	0.338				

Data presented as mean \pm SE

† Based on independent t-test.

§ Based on linear regression (the included variables were: basic value of dependent variable, treatment type, age and BMI)

P value within groups based on paired t-test

*statistically significant.

SIRT1: sirtuin-1, PGC1 α : Peroxisome proliferator activated receptor γ coactivator 1 α

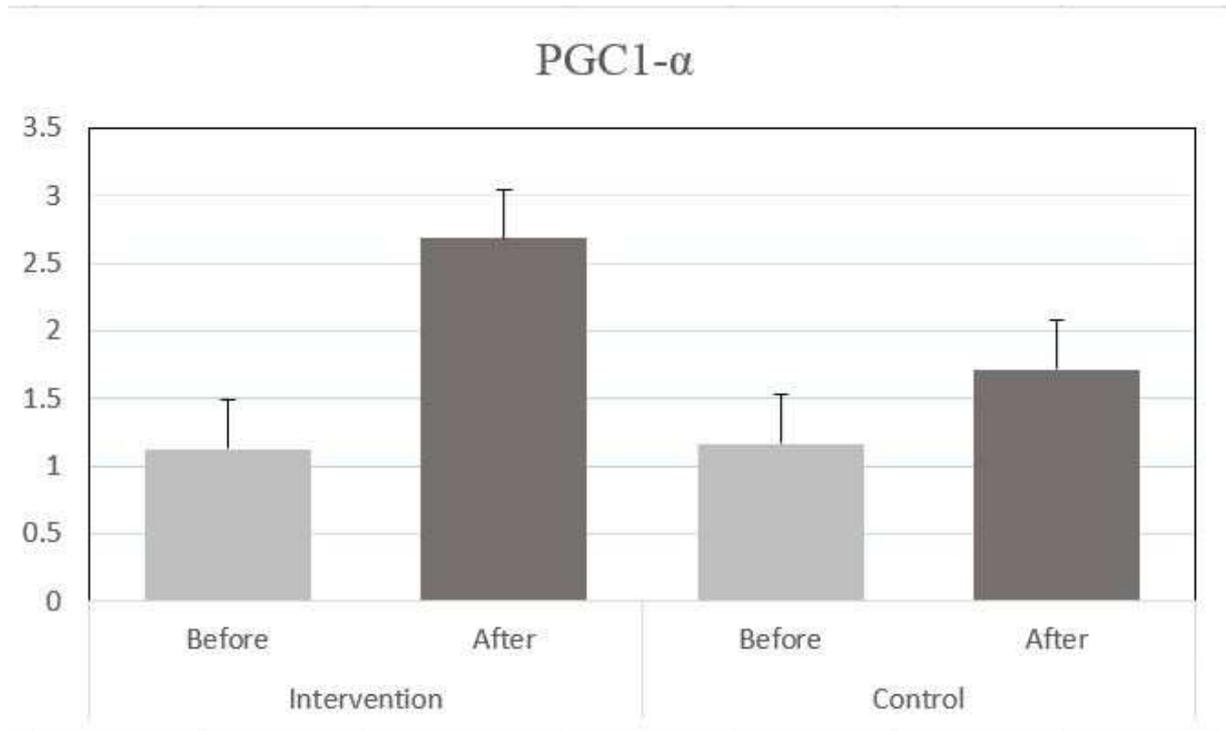


Figure 2- Effect of curcumin on gene expression of Peroxisome proliferator activated receptor γ coactivator 1 α (PGC1- α) (data presented as $2^{-\Delta\Delta ct}$)

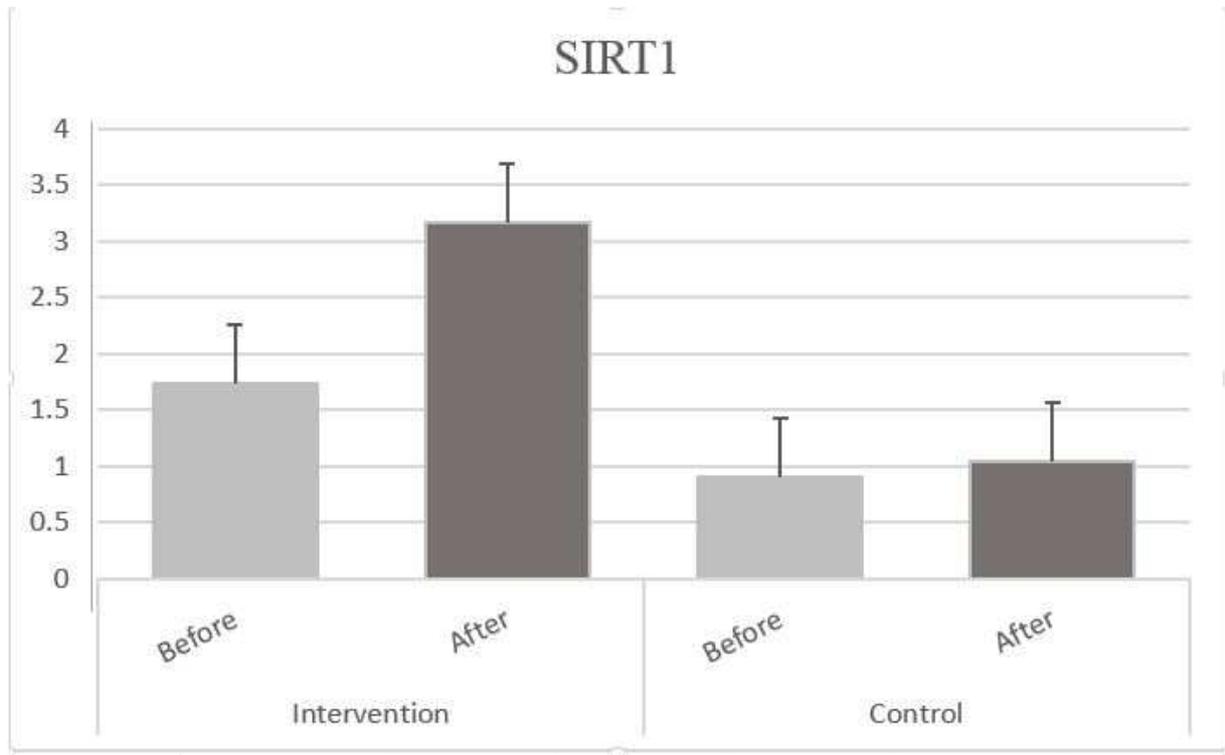


Figure 3- Effect of curcumin on gene expression of Sirtuin-1 (SIRT1) (data presented as $2^{-\Delta\Delta C_t}$)



Iran University of Medical Sciences

Research Ethics Certificate

Approval ID:	IR.IUMS.REC.1397.690	Approval Date:	2018-10-28
Evaluated by:	Iran University of Medical Sciences		
Status:	Approved		
Approval Statement:	<p>The project was found to be in accordance to the ethical principles and the national norms and standards for conducting Medical Research in Iran.</p> <p>Notice:</p> <ol style="list-style-type: none"> 1. Although the proposal has been approved by the research ethics committee, meeting the professional and legal requirements is the sole responsibility of the PI and other project collaborators. 2. This certificate is reliant on the proposal/documents received by this committee on 2018-10-28. The committee must be notified by the PI as soon as the proposal/documents are modified. 		
Proposal Title:	The effect of curcumin supplementation on PGC1 and SIRT1 gene expression in polycystic ovary syndromePCOS woman		
Principal Investigator:	Name: Farzad Shidfar Email: shidfar.f@iums.ac.ir		

Dr. Masood Naseripour
 Director of University/Regional Research Ethics
 Committee
 Iran University of Medical Sciences

Dr. Seyed Kazem Malakouti
 Secretary of University/Regional Research Ethics
 Committee
 Iran University of Medical Sciences

Permission letter from ethics committee